

Induced polyploidy in *Betula*

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TIIVISTELMÄ: INDUSOITU POLYPLOIDIA BETULA-SUVUSSA

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Seeds of ten different *Betula* species were treated with colchicine solution during germination, to induce duplication of the chromosome set. The species included in the study were *B. pendula*, *B. pubescens*, *B. papyrifera subcordata*, *B. papyrifera papyrifera*, *B. papyrifera occidentalis*, *B. populifolia*, *B. alleghaniensis*, *B. nigra*, *B. glandulosa* and *B. nana*. The total number of individually labelled, colchicine-treated trees was 1550. Colchicine treatment induced changes in morphological features, especially in the leaves. These features proved to be good indicators of polyploidization. The experiments produced 687 polyploid trees, 287 of which are still alive. The polyploid *Betula* trees offer possibilities of studying the significance of the genome dosage for the growth, breeding, adaptability and evolution of *Betula*.

Kymmenen eri koivulajin siemeniä käsiteltiin kolkisiiniliuoksella itämisen aikana tavoitteena kromosomiston kertautuminen. Käsitellyt lajit olivat *B. pendula*, *B. pubescens*, *B. papyrifera papyrifera*, *B. papyrifera subcordata*, *B. papyrifera occidentalis*, *B. populifolia*, *B. alleghaniensis*, *B. nigra*, *B. glandulosa* ja *B. nana*. Yksilökohtaisesti merkittyjen, kolkisiiniliuoksella käsiteltyjen puiden lukumäärä oli 1550. Kolkisiinikäsittely aiheutti muutoksia morfologisissa ominaisuuksissa, erityisesti lehdissä. Nämä ominaisuudet osoittautuivat hyviksi polyploidisaation osoittajiksi. Kokeet tuottivat 687 polyploidia puuta, joista on vielä elossa 287. Polyploideja koivuja voidaan hyödyntää koivun jalostuksessa sekä tutkittaessa ploidiatason merkitystä kasvuun, sopeutuneisuuteen ja koivujen evoluutioon.

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1. Introduction

Natural polyploidy is very frequent in the genus *Betula*, which comprises approximately 40 different species (e.g. Winkler 1904, McCown 1989). The number of ploidy levels differs between the four subsections of the genus. The subsections *Costatae* and *Albae* are the most variable ones: *Costatae* has diploid ($2X=28$),

tetraploid ($4X=56$) and hexaploid ($6X=84$) *Betula* spp., and, in addition to these three ploidy levels, *Albae* also has a pentaploid ($5X=70$) species. *Nanae* has both diploid and tetraploid species, *Acuminatae* only diploid ones.

The great natural variation in the number of chromosome sets in *Betula* and the great

adaptability of this genus to different environments, created the starting point for this research. The possibility of using induced polyploidy to speed up the natural evolution and breeding of *Betula* arose in the 1960's.

The purpose of this research is to study how resistant or susceptible the species of the widespread polyploid genus *Betula* are to artificial manipulation of the genome. A further aim is to create large, new materials for basic and breeding studies of *Betula* spp. *In vitro* methods allow efficient use of the present, artificially polyploidized trees in studying genetic identity between micropropagules and the mother tree and among progenies. Being ontogenetically

2. Materials and methods

The materials of this research represent three of the four subsections into which *Betula* is divided: *Albae*, *Costatae* and *Nanae* (Table 1). The different *Betula* species used, and their seed origin codes are presented in Table 1. A detailed summary of the seed origins was presented by Vaarama and Valanne (1969) and Valanne (1972).

The polyploidization experiments were performed by treating seeds with colchicine during germination in 1962–1967. Seeds were either kept in colchicine solution for the whole germination period or alternately in colchicine solution and distilled water. The different colchicine treatments and their code numbers as well as the numbers of individually labelled seedlings produced and

very old due to the aging effect of colchicine, the colchicoid trees are convenient explant trees in studying rejuvenation through meristem culture. Developing *in vitro* culture methods for polyploid birches is also studied in this research programme.

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their total numbers are shown in Table 1.

The chromosome determinations were done either on squashed root tips of rooted leaf petioles (Valanne 1978) or on flushing leaves, according to a double-staining method (Hömmö and Särkilähti 1986).

The young colchicine-treated birches were transferred to their final growing sites at the age of 4–5 years in 1965 and 1967–69. The locations of the different colchicine treated trees are on the island of Seili (60°12'N, 21°55'E), at the Experimental Station of the Finnish Forest Research Institute, Punkaharju (61°43'N, 29°25'E), in the Botanical Garden of the University of Turku, Ruissalo (60°26'N, 22°10'E), and at Päilähti by Orivesi (61°37'N, 24°29'E; Figs. 1–2).

3. Results

A rough estimate of the number of suspected polyploids was obtained using the colchicine-induced morphological features described by Valanne (1972) as an indication of polyploidization. These features are a robust habit, ramification of the trunk and especially aberrant leaf morphology (Fig. 2).

On the basis of the colchicine-induced changes in leaf morphology, the trees were divided into three different morphological groups: no morphological differences compared with normal trees, slight colchicine morphology (++) and pronounced colchicine morphology (+++).

Table 1. *Betula* materials and treatments used in the colchicine experiments in 1962–1967. (C = colchicine solution, W = distilled water)

Subsection Species	Seed origin numbers	Treatments and their codes ¹	Individual tree numbers	Total number of trees
<i>Albae</i> <i>B. pentula</i>	1, 3–4, 8–9, 12–13, 16–18	4.–54.–62.*	98–853	755
	6	2.–63: 2x(1C+1W)+1C+7W 0.06% C	860–867	8
	15	46.–63:	2037–2048	12
	15	1.–64: 10 000r + 3x(2C+1W)+2C+3W 0.06% C	2213–2367	155
	15	2.–64: 15 000r + "	2368–2397	30
	6	3.–64: 2x(2C+1W)+2C+6W 0.06% C	2398–2410	13
<i>B. pubescens</i>	19	4.–64: 17 days in 0.1% C	2411–2458	48
	19	4.–64: " 0.05% C	2459–2512	54
	19	5.–64: 5x(1C+1W) 0.1% C	2514–2550	37
	19	6.–64: 17 days in 0.04–0.05% C	2576–2600	24
<i>B. papyrifera subcordata</i> <i>B. papyrifera papyrifera</i> <i>B. papyrifera occidentalis</i> <i>B. populifolia</i>	43	13.–64: 5C+8W 0.06% C	2633–2651	19
	42	16.–64: 3C+2W+2C+8W 0.06% C	2835–2988	154
	44	14.–64: 2C+3W+5C 0.06% C	2652–2776	125
	46	19.–65: 10 000r + 0.06% C 4 days	3410–3442	33
<i>Costatae</i> <i>B. alleghaniensis</i> <i>B. nigra</i>	46	21.–65: 15 000r + 0.06% C 4 days	3489–3511	23
	41a 47	15.–64: 2C+4W+7C 0.06% C 5a.–67: 5C+0W 0.1% C	2777–2834 **	58
<i>Nanae</i> <i>B. glandulosa</i> <i>B. nana</i>	30			
	63			

¹ Seeds were either germinated in colchicine or kept alternately in colchicine solution and distilled water. The explanation to 2x(1C+1W)+1C+7W: seeds were first kept in colchicine solution for one day, then in distilled water for another day. This was repeated. Thereafter the seeds were again kept in colchicine for one day, and finally in distilled water for seven days.

* many treatments (Vaarama and Valanne 1969: 16–17)

** 56 saplings, all died in winters 1969–1970 and 1970–1971

*** no surviving seedlings

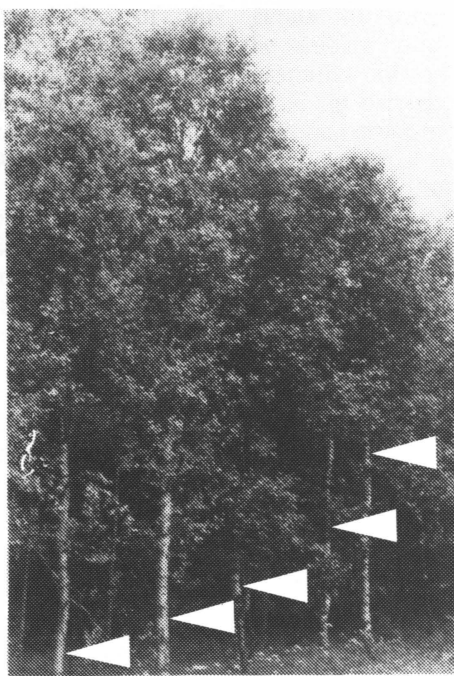


Fig. 1. Polyploid *Betula pendula* and *B. pubescens* materials in Päilahti, Orivesi in 1988. Trees marked with arrowheads from right to left:

<i>B. pendula</i> no.	2382	2n=56,	irradiation-
			mutant,
"	862	2n=56,	
"	2315	2n=56(60),	
<i>B. pubescens</i> no.	2440	2n=70-84 and	
"	2485	2n=98.	

This division was used when a sample of adult *B. pendula* Roth and *B. pubescens* Ehrh. trees was chosen for chromosome determinations in 1988 (Tables 3 and 4, Figs. 1-2).

The numbers of induced polyploid trees of different *Betula* confirmed during the juvenile and adult phases (1969 and 1987) are summarized in Table 2. Polyploids were recorded in all the investigated species except *B. nana* L. and *B. glandulosa* Michx.. The total number of treated and individually labelled trees in the inventory of 1969 was 1550. The total number of polyploids confirmed during the juvenile phase was 687 (Vaarama and Valanne 1969) and approximately half of these are still alive.

Tables 3 and 4 exemplify the great variability in chromosome numbers of the

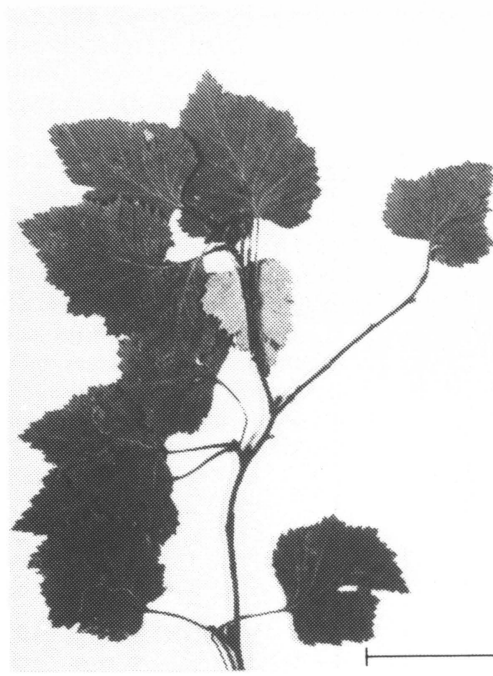


Fig. 2. Leaf specimen of *B. pendula* tree no. 2382. Scale 5 cm.

adult colchicine-induced polyploids of both *B. pendula* and *B. pubescens*. The abundance of aneuploids is also conspicuous in both species. In *B. pendula*, colchicine seems to induce autotetraploids and aneuploids in the ratio of 2:1. In *B. pubescens*, however, half of the induced polyploids have the chromosome number 98. Fig. 1 shows polyploid *B. pendula* and *B. pubescens* trees in Päilahti, Orivesi in 1988.

In addition to the C_0 generation (colchicine-treated trees), our materials include C_1 progenies of colchicine-polyploid individuals of three species: *B. papyrifera* Marsh. (tree no. 2867.16.-64, $2n=140$), *B. pendula* (tree no. 314.29.-62, $2n=56$), and *B. pubescens* (tree no. 2574.6.-64, $2n=112$). The analysis of these open-pollinated 9-12 years old progenies is in progress (Vataja, unpublished). The C_0 *B. papyrifera* tree produced hexaploid seedlings ($2n=84$), the *B. pendula* triploid ($2n=42$) and the *B. pubescens* tetraploid and pentaploid ($2n=56, 70$) seedlings (Särkilahti, unpublished).

Table 2. The numbers of polyploid *Betula* trees in 1969 and 1987 at different sites.

Species	Treatment no.	Numbers of polyploids analyzed in 1969 ¹	Site							
			Seili ¹		Punkaharju ¹		Orivesi			
			1969	1987	1969	1987	1969	1987		
<i>B. pendula</i>	4.-54.-62	188	137	-			41	22	10	3
	2.-63	6			1	1			4	2
	46.-63	9			4	2			2	1
	1.-64	136			116	72	13	8	2	1
	2.-64	30			26	15	3	3	1	1
	3.-64	13			13	10				
<i>B. pubescens</i>	4.-64	77			46	33	5	3	10	5
	5.-64	36			22	18	5	4	9	3
	6.-64	22			10	9	10	5		
<i>B. papyrifera subcordata</i>	13.-64	12			6	-	4	4	2	0
<i>B. papyrifera papyrifera</i>	16.-64	86			49	-	27	9	10	6
<i>B. papyrifera occidentalis</i>	14.-64	58			18	-	30	18	10	6
<i>B. populifolia</i>	19.-65	-					18	6	15	3
	21.-65	-					13	8	10	1
<i>B. alleghaniensis</i>	15.-64	8					6	4	2	1
<i>B. nigra</i>	5a.-67	6					6	0		

¹ Dash means lack of observation

4. Discussion

Comparison of the leaf morphology with the cell size in different tissues and the chromosome numbers (Pieninkeroinen and Valanne 1989, Särkilahti and Valanne 1990) showed that identification of adult polyploid trees is possible visually on the basis of the leaf morphology in the *B. pendula*, *B. pubescens* and *B. papyrifera* complex. The other species do not show such strong colchicine-induced changes in leaf morphology.

The characteristics that are most indicative of polyploidization are larger leaf blades, thicker leaf petioles and a rougher network structure on the abaxial leaf epidermis than in normal trees. These are all consequences of the enlarged cell size in the leaves (Pieninkeroinen and Valanne 1989, Särkilahti and Valanne 1990).

The C_1 progenies are characterized by severe growth retardation in all growth phases. This result is in disagreement with that of Johnsson (1956), whose C_1 progenies

showed only minor growth retardation. In general, the flowering ability of C_0 trees seems to vary greatly within the genotype, as can be concluded from our results and from previous experiments (Johnsson 1956, Eifler 1967).

The most effective method of polyploidization has generally proved to be colchicine treatment during seed germination. Compared with the large number of induced polyploids in several *Betula* species presented in this report, few references to successful polyploidization in *Betula* species can be found in the literature. The previous experiments comprise only three species: *B. pendula*, *B. pubescens* and *B. latifolia* Komar. (Johnsson and Eklundh 1940, Schröck 1951, Eifler 1955, Hyun and Kim 1963). In their experiments, colchicine was given in overdoses: high concentrations (e.g. 0.2%) and long treatment times were used. This resulted in a low number of polyploids, evidently due to the large

Table 3. Chromosome numbers of colchicine-induced polyploids of *B. pendula* trees.

Treatment no.	Tree no.	Morphological groups ¹	Chromosome numbers (2n)	See figure
48.-62	745	+++	56	
51.-62	770	+++	56	
2.-63	862	+++	56	1
"	865	+++	56	
46.-63	2040	++	56	
1.-64	2213	++	56	
"	2229	++	56	
"	2245	++	56	
"	2248	++	56	
"	2265	++	56	
"	2266	++	28, 29, 42, 48, 50	
"	2267	++	56	
"	2283	++	56, ca.60, ca.84, ca.98	
"	2293	++	56	
"	2303	++	56	
"	2315	+++	56 (60)	1
"	2347	++	28, 42, 56, 70, 30, 34, 37	
"	2361	++	56	
2.-64	2372	++	56	
"	2382	+++	42, 56	1 and 2
"	2386	++	56	
"	2388	+++	56	
"	2389	++	56, ca.60	
3.-64	2401	+++	28, 56	

¹ Key: ++ = slight colchicine morphology of leaves
+++ = pronounced colchicine morphology of leaves

proportion of undeveloped seedlings and the high mortality. Johnsson and Eklundh (1940) achieved polyploidization in eight *B. pendula* individuals and two *B. pubescens*. Three of their polyploidized *B. pendula* trees started flowering at the age of 12–13 years (Johnsson 1956). The colchicine experiments performed by Schröck (1951) with *B. pendula* produced only six polyploid individuals (Eifler 1955), which were examined at the age of 19 years, at which time none of the trees had flowered (Eifler 1967). Two polyploid individuals of *B. latifolia* were obtained by Hyun and Kim (1963).

The most effective technique for induction of polyploidization in *Betula* species was developed in the course of the experiments in 1962–1966, as shown by the great number of induced polyploid individuals in several

Betula species (Tables 2–4). The technique of holding seeds alternately in colchicine solution and distilled water, using a low colchicine concentration (0.06%), resulted in a large number of polyploids. This technique did not arrest the cell mitosis cycle due to a too high polyploidization rate, but affected only one mitosis cycle, resulting in continuing but decelerated mitosis. These results suggest that the genomes of the various *Betula* species are far from conservative and can be manipulated artificially to produce genotypes with higher ploidy levels, unlike those of coniferous tree species, which seem to become inviable beyond their natural ploidy level (e.g. Johnsson 1975, Illies 1976). In general, deciduous tree species are younger from the evolutionary point of view than coniferous

Table 4. Chromosome numbers of colchicine-induced polyploids of *B. pubescens* trees.

Treatment no.	Tree no.	Morphological groups ¹	Chromosome numbers (2n)	See figure
4.-64	2424	++	ca.98	
"	2426	++	ca.70–98	
"	2428	++	98	
"	2433	++	ca.112	
"	2435	++	ca.98	
"	2439	++	ca.98	
"	2440	+++	70–84–98	1
"	2452	++	98, 70, 56	
"	2461	++	ca.98	
"	2463	+++	98	
"	2464	++	ca.84–98	
"	2465	++	ca.98	
"	2466	++	ca.98	
"	2468	++	ca.98	
"	2473	+++	ca.56, ca.70–84	
"	2475	+++	84–98	
"	2476	++	ca.98	
"	2477	++	ca.98	
"	2480	++	ca.98	
"	2481	+++	80–98	
"	2485	+++	98 (112 ?)	1
"	2490	++	ca.84–98	
"	2494	++	ca.98	
"	2496	++	ca.98, ca.100	
"	2497	++	ca.98, 112	
"	2498	++	ca.90	
"	2504	++	ca.112	
5.-64	2514	++	90–100	
"	2515	++	ca.98	
"	2516	++	ca.98	
"	2520	++	ca.98	
"	2521	++	ca.112	
"	2529	++	98–112	
"	2535	++	ca.98	
"	2538	++	ca.98, 104 ?	
"	2542	+++	98	
"	2544	+++	98, 112	
"	2548	++	ca.98	
6.-64	2571	++	84–100	
"	2579	++	ca.98–101	

¹ Key: ++ = slight colchicine morphology of leaves
+++ = pronounced colchicine morphology of leaves

trees. This explains the polyploid nature and the occurrence of various ploidy levels, e.g. in *Betula*. Evolution is still proceeding in *Betula*, as shown by the frequent cases of hybridization and introgression at the northern tree limit in Fennoscandia (Sulkinoja 1981, 1983, Kallio et al. 1983).

The birch materials presented in this paper offer possibilities of studying the significance of the genome dosage for the growth, breeding, adaptability and evolution of *Betula*. A large polyploid series is available, consisting of both natural and induced polyploids. Ploidy levels ranging from diploid to dodecaploid (2X, 3X, 4X, 5X, 6X, 8X, 10X and 12X) are represented in these materials. In the colchicine experiments reported here, polyploids were achieved in all the three sections used, except *Nanae*, which did not produce any viable seedlings from colchicine treatment at all. *Nanae* birches evidently lose their viability at other ploidy levels than their natural one.

Thus, we have large autopolyploid materials of different *Betula*, which can be maintained through micropropagation methods. Some of our colchicine-treated and polyploid individuals have already been cultured as sterile shoots, which have also produced micropropagated plantlets (Särkilahti 1988, 1989). Clonal materials allow testing of genetic identity both between the parent tree and its clonal progeny, and within a clonal progeny (e.g. McCown 1989). A significant question in the production of micropropagated forest trees is whether micropropagation results in true rejuvenation (Hackett 1985, Francllet et al. 1987) or merely an epigenetic change (Varga et al. 1988, Karp 1989). The effects of topophysis and cyclophysis in the meristems of adult trees (Evers 1987) may not be truly overcome by *in vitro* culture, but only delayed due to environmental factors in the culture procedures. This would produce rejuvenated plantlets and juvenile trees which would switch to the adult phase much earlier than trees of the same age originating from seed. The results on micropropagation of an adult irradiation-mutant and colchitetraploid of *B. pendula* suggest that there is no true rejuvenation induced during the culture of the meristems (E. Särkilahti, unpublished).

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Total of 27 references